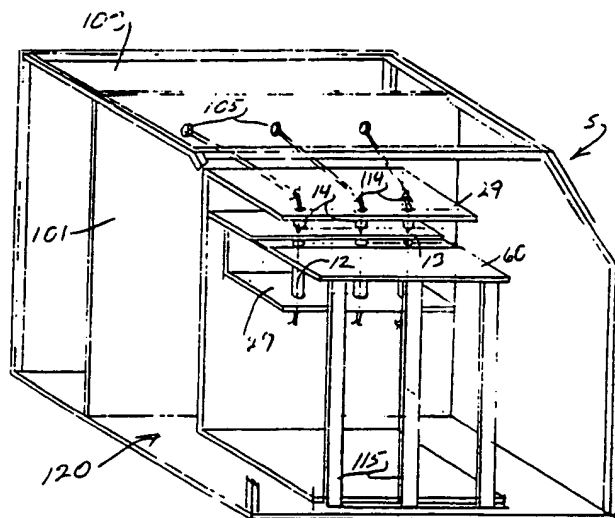




INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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(21) International Application Number: PCT/US90/05238 (22) International Filing Date: 19 September 1990 (19.09.90) (30) Priority data: 409,108 19 September 1989 (19.09.89) US (71) Applicant: PARK PHARMACEUTICALS INC. [US/US]; 11880 Bird Road, Suite 101, Miami, FL 33175 (US). (72) Inventors: ANDREOTTI, Peter, E. ; 3940 N.W. 75 Ter., Lauderhill, FL 33319 (US). MORSE, Irwin, S. ; 151 S.E. 15 Road, Miami, FL 33129 (US). THORNTWHAITE, Jerry, T. ; 7880 S.W. 139 Ter., Miami, FL 33158 (US). HEIMER, Malcolm, L. ; 11275 S.W. 128 Court, Miami, FL 33186 (US). SALINGER, Jorge, D. ; 6706 S.W. 152 Court, Miami, FL 33193 (US). SOBODOWSKI, Joseph, J. ; 10605 S.W. 135 Court, Miami, FL 33186 (US).	(74) Agents: WOLFFE, Franklin, D. et al.; Fidelman & Wolffe, 1233 20th Street, N.W., P.O. Box 18218, Washington, DC 20036-8218 (US). (81) Designated States: AT (European patent), BE (European patent), CH (European patent), DE (European patent)*, DK (European patent), ES (European patent), FR (Eu- ropean patent), GB (European patent), IT (European patent), JP, LU (European patent), NL (European pa- tent), SE (European patent). Published With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.	

(54) Title: LUMINOMETER



(57) Abstract

A luminometer (5) for assaying bioluminescence and chemiluminescence reactions characterized by sample wells (30) having transparent bottom walls, a light detector (12) positioned beneath the sample wells and a reagent injector above the wells. A sample holder (16) desirably the size of a microtiter plate is comprised of an opaque body member formed with a multiplicity of pass through cylindrical apertures plugged by cup shaped sample wells with their transparent bottom walls recessed in the apertures. Optionally the side walls of the sample wells have a reflective coating. This invention relates to a novel luminometer and in particular to a luminometer which employs luminescence to perform a series of analytical tests rapidly.

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LUMINOMETER

BACKGROUND OF THE INVENTION

For some years prior to the date hereof the art has appreciated that luminescence reactions, particularly bioluminescence and chemiluminescence reactions can be basis for extraordinarily sensitive analytical assays or tests. Such assays can, for example, combine the specificity of an immunoassay with the detection limits for measurements of light. For instance, luminescent immunoassays are comparable in sensitivity to radioimmunoassays and offer the significant advantages of freedom from radioactive reagents.

However, as of date hereof the potential for rapid accurate assays offered by bioluminescence and chemiluminescence reactions has not been achieved. To some extent, perhaps to a great extent, the analytical luminometer instruments offered to the art heretofore are ill-suited to rapid processing of numerous samples in luminescence reactions assays. Certainly, they have not met with widespread acceptance. This invention relates an improved luminometer instrument for conduct of luminescence, (i.e., bioluminescence or chemiluminescence) assays.

RATIONALE OF THE INVENTION

An attribute desired as much in a luminometer as in any analytical instrument is the capability to analyze a great number of similar samples one by one, quickly and accurately. Improved sensitivity is, of course, desired.

These objectives have been achieved by the luminometer of this invention.

In the luminometer of this invention a large number of samples placed in a sample holder that comprises a modified microtiter plate are introduced into the luminometer instrument at one time, then measured, one by one in some

sequence until the entire batch of samples on the sample holder has been analyzed.

The well known microtiter plate which typically contains wells in 8 rows and 12 columns, namely, 96 wells, each of about 0.25-0.30 ml capacity, is a conveniently sized sample holder. Moreover, microtiter plates are small enough so that several, e.g., three modified microtiter plates, can fit into a luminometer of convenient size for an analytical laboratory. Thus, 96 samples in triplicate or the near to 300 individual samples in the wells of three modified microtiter plates sample holders may be assayed in a single batch.

It goes without saying that the large number of samples that may be placed into one or more of the sample holder plates will not be assayed simultaneously (by some luminescence reaction). They may, in fact, be assayed one at a time; conceivably only one light detector and one inlet tube for adding necessary reagent(s) to each sample need be in the luminometer. Preferred practice of this invention, however, is to provide a light detector and reagent feed tube(s) for each sample holder plate. Preferred is three plates, three light detectors, etc., in the luminometer. The plates are spaced apart so measurements made luminescence on one plate will not interfere with measurements on the other plates.

Accordingly, necessary components in the luminometer of this invention are a movable table on which the sample holder plate(s) are removably mounted, an X, Y movement mechanism which generates closely controlled movement of the table repetitively accurate to about 0.0015 inch or better, at least one sample holder plate, and a light detector adjacent to and associated with the sample holder plate. Each sample (well) in the sample holder plate is brought into line of sight registry with the light detector

in some predetermined sequence. Also necessary is one or more inlets in an appropriate registry with the light detector for luminescence assay to introduce reagent(s) needed for the luminescence reaction into each sample in turn.

Luminometers systems such as are discussed above have been suggested to the art as, for example, by U.S. patent 4,772,453 and by EPO publication 0025350 published March 18, 1981.

The present invention is directed to an improved luminometer system.

THE INVENTION

For further understanding of the present invention reference is made to the attached drawing illustrating a preferred mode of the invention and wherein:

Fig. 1 is a diagrammatic view of the luminometer housing;

Fig. 2 is a diagrammatic view of the luminometer with the housing and moving parts removed showing the mountings for the light detector and reagent injector;

Fig. 3 is a diagrammatic view of the luminometer with the housing and some parts removed, showing the X-Y mechanism and movable table;

Fig. 4 is a side section through the luminometer;

Fig. 5 is a front view section through the luminometer at the location of line 5-5 on Figure 4;

Fig. 6 is an enlarged partial section corresponding to Figure 5, showing a sample well on the sample holder plate.

Referring now to the drawing and, in particular to Fig. 4 and Fig. 5 whereon is shown luminometer 5 containing the components which comprise the luminescence assay system of this invention, namely, sample holder 16, a light sensor 12 beneath sample holder 16 on a line of sight to one sample at a time, the sensor in the instance of this invention preferably being a state of the art photo multiplier tube,

e.g., Hamamatsu R647-04 and an injector 14 above the sample holder for introducing the reagent(s) which generate(s) the desired luminescence reaction in the liquid sample. Needed also is a read out means (not shown) that reports the level
5 and/or quantity of luminescence measured by photomultiplier tube 12. Read out means for photomultipliers are state-of-the-art systems.

In the luminometer read out employed as of the date hereof, the signal pulses from the photomultiplier tube
10 have been amplified, then passed through a discriminator circuit (to distinguish noise from signal) and thereafter counted over the preselected assay test time interval. After subtraction of the background count levels, the net pulse count (or some fraction or multiple thereof as
15 appropriate) was reported as the raw data of the luminescence assay, usually to a computer for a later conversion into a (printed out) spread sheet report on all of the samples in a batch. The circuitry and other components for this and like read out means are known in
20 the art forming, per se, no part of this invention.

The luminometer of the present invention provides for assaying, one by one, the numerous samples, e.g., 96 that can be placed in the multiplicity of sample wells 30 in sample holder plate 16, (see Fig. 6) preferably in a batch
25 of three such sample holder plates 16 as is illustrated in Figure 5.

Referring now to Figures 2, 3 and 4, it may be seen there that the sample holder plate(s) 16 are removably mounted on a table 110 that is movable relative to the stationary
30 photo multiplier tube(s) 12 positioned below table 110 and the injector 14 positioned above Table 110. An X-Y drive mechanism 18, of which many are known to the art and, therefore, is not illustrated, being shown fragmentarily in Figures 3 and 4 shifts table 110 relative to the

photomultiplier tube (PMT) 12 and injector 14 so that each sample well on the sample holder plate(s) will come into registry with the PMT and also with injector 14 one by one. To repeat, in the luminometer of this invention PMT 12 and
5 injector 14 are stationary.

Although the foregoing discussion speaks of the table being movable, and PMT and injector being stationary, it should be appreciated that the movable/stationary relationship applies only with regard to lateral movement
10 (e.g., in the plane of the table) by the table. Indeed, vertical (upward) movement by injector 14 and/or vertical (downward) movement by PMT 12 may be desired to provide room for insertion of sample holder plate 165 on Table 110.

15 As may be seen best in Figures 3 and 4, Table 110 forms part of a movable chassis 112 that is linked to the X-Y mechanism 18 for controlled movement by the X-Y mechanism. Movable chassis 112 is inside of a stationary cage 115, whereon is mounted the injectors 14 and the PMT 12 above
20 and below Table 110 respectively. Injectors 14 are shown as mounted on an upper shelf 29 on cage 115. The PMT 12 are shown as seated on a lower shelf 27. Between the shelves 27, 29, and beneath Table 110 is a baffle plate 60. The baffle plate 60 is shown as extending to legs 117,
25 which supports may be made part of the front face of the housing 100. Legs 117 are removed for access to X-Y mechanism 18 and to chassis 112 when, as and if maintenance becomes necessary. Conveniently, the luminometer 5 is constructed with a frame 6 around which the sheet metal
30 housing 100 is draped.

Reverting now to Figures 1-3 of the drawing it may be seen there that access to the inside of luminometer housing 100 for placement and removal of the sample holder plates therein, is through pivotal movement of a hatch 102 from a

light tight closed position to an open position. When hatch 102 is open, sample holder plates each with a multiplicity of samples in the sample wells 30 may be placed (by hand) into the three recesses 104, 106, 108 provided in table 110 for this purpose and, of course, already assayed plates in any of the recesses are removed then.

Desirably an internal well 101 is provided inside housing 100, generating thereby a front compartment 120 containing chassis 112, etc., and rear compartment 103 wherein may be placed all ancillary equipment, e.g., electrical controls, and reagent supply vessels for injectors 14, etc. A reagent supply tube 114 passes from behind internal wall 101 through a port 105 to the injector 14.

In the luminometer of this invention, the reagent injector 14 normally is positioned above the microtiter plate in line with PMT 12. If desired for some luminescence assays injector 14 may be offset (e.g., by one sample). The offset for reagent injector 14 is contemplated for instances where a delay between addition of the reagent and the assay measurement of the resulting luminescence reaction is desired. The offset then is placement of injector 14 at the sample that next will be in line with the PMT or the next succeeding sample, as appropriate. All such locations for injector 14 are considered to be in registry with the PMT. In the mode illustrated, only one reagent injector 14 is present for each sample holder and that injector is in line with the PMT 12 as is indicated on Figures 2 and 4. For assays where more than one reagent may be needed, multiple injector nozzles, as necessary will, of course, be present and their placement will be determined by the intended luminescence reaction.

Referring now to Figures 5 and 6, it may be seen there that injector 14 comprises an assembly wherein a nozzle tip 13 is mounted on the nozzle mounting fixture 17 located on upper shelf 29. Fixture 17 comprises a plug 22 through which the nozzle tip 13 passes and within which the nozzle tip 13 is firmly retained. In turn, the plug 22 is secured in a threaded nut 24. Nut 24 is threaded to upper and lower gaskets 26 and 28 which together lock gaskets and plug to a depending lip or flange on shelf 29.

Each sample well 30 on sample holder plate 16 is provided with a transparent bottom wall 31 so that light generated by the luminescence reaction can pass through (the transparent bottom wall 31) to strike the photo sensitive face of PMT 12. Light from all other sources is avoided.

As will be explained hereinafter, several important geometric features are provided in the luminometer of this invention to eliminate impingement of unintended light upon the photo sensitive face of PMT 12.

As already has been pointed out, each sample 10 is disposed inside of a well 30, one of many such wells, e.g., ninety-six in the sample holder plate 16. To repeat, the sample holder plate 16 may be considered to be modified microtiter plate. Like typical microtiter plates the sample holder plate 16 employed with the luminometer of this invention contains a dozen columns numbered 1-12 and 8 rows numbered A-H for a total of ninety-six wells (all as is present in known in the art microtiter plates). The outside dimensions of the plate, the well diameter and well center to center spacing may be the same as in conventional microtiter plates, e.g., the clear molded polystyrene Cell WellTM plate from Corning Glass Works. This mimicry is advantageous since considerable ancillary equipment has been engineered for use with microtiter plates and, therefore, reliable pipetting and control equipment is

commercially available for use with the sample holder plates of this invention (e.g., Cavity Instruments) to add, remove, or transfer solution(s) to and/or from up to 12 wells simultaneously on demand. Thus, conceivably the
5 (modified microtiter plate) sample holder plate of this invention could be employed as a microtiter plate, but, of course, the known in the art microtiter plates are not adapted to practice of this invention.

The body 35 of sample holder plate 16 should be opaque,
10 preferably being black. The wells 30 comprise cup shaped inserts with transparent bottoms which are pressed into pass through cylindrical apertures 39 in body 35. The entire well 30 may be transparent or alternatively the side wall of well 30 may be light reflective. However, it is
15 necessary to practice of this invention for the bottom wall 31 of each well 30 to be transparent.

A feature of importance to practice of this invention is that transparent bottom wall 31 of each well 30 is spaced inward from the (planar) lower surface 38 of the sample
20 holder plate body 35. A small but distinct (cylindrical) recess 40 has thereby been created beneath each well 30.

Thus, the drawing, notably Figures 5 and 6 illustrates a preferred mode wherein the sample holder plate 16 comprises a black molded body 35 member having therein cylindrical
25 pass through apertures 39 set in the column and row array present as in microtiter plates. Each aperture 39 is plugged by cup shaped well 30 with transparent walls. The well 30 seats in the aperture 39 with its top opening flush to the upper surface of plate body 35 but its transparent
30 planar bottom wall 31 recessed above the bottom surface 38 of the molded plate body 35.

The absolute dimensions of the sample holder plate 16, of the well 30 and of the recess 40 are not critical to practice the invention, although, to repeat, it is

advantageous for plate 16 to have the same dimensions as a typical microtiter plate. A sample volume of about 0.25 ml is convenient for luminescence assay purposes. Exemplary dimensions for plate 16 are 1.5 cm for the depth of the plate body 35, about 1 mm for the (depth of) recess 40, an internal diameter of about 6 mm, and a depth of 1.3 cm for well 30. More generally, the depth of recess 40 may vary from 0.5-1.0 mm.

Known in the art photo multiplier tube(s) (PMT) 12, e.g., Hamamatsu R647-04 and read out means such as has been described above are employed in the luminometer 5. Front compartment 120 in housing 100 wherein the PMT 12 are disposed is a light tight compartment. Hatch or lid 102 must fit securely. Thus, PMT 12 is exposed to light only through viewing aperture(s) 65 in the baffle plate 60 provided for this purpose as may be seen in Fig. 5, and Fig. 6.

The PMT 12 is disposed beneath the upper surface of (stationary) baffle plate 60, desirably inside of a cylindrical cutaway 62. The baffle plate 60 is opaque and desirably is black.

An annular focussing collar 64 for the PMT face is disposed in cutaway 62 (above the PMT) face leaving an iris passageway 65 that constitutes the viewing aperture through which light from sample 10 strikes the photo sensitive face of PMT 12. Preferably, the diameter of iris 65 is somewhat less than the diameter of sample well 30, being for example from 0.5-3.0 mm less in diameter. The focussing collar 64 is, of course, opaque, and preferably is black.

It should be appreciated that wells 30 are small and are close together, for example, a particular well 30 being illustrated in Figure 6 at two-three times full size; for clarity of illustration the next adjacent wells to that

illustrated in full are shown by Figure 6 only in shadow form.

Careful construction of the luminometer as a whole and of the component parts therein is needed. For one thing a reasonable clearance above and below table 110 (and the sample holder plates thereon) must be provided to allow for the movement of table 110 that brings each well of the microtiter plate into registry with PMT 12 and injector 14 in the desired succession but more than enough clearance is not desirable.

As has already been pointed out, the injector 14 must be in its own registry with PMT 12, e.g., injector tip 13 in line of sight with PMT 12 so as to feed reagent directly into the sample 10 in the particular well 30 on line of sight to PMT 12, for readings on rapid onset reactions.

The X-Y movement mechanism must move the table 110 exactly the distance center to center between (adjacent) sample wells, to an accuracy of ± 0.0015 or about 0.04 mm. A like level of uniformity in the sample holder plates is important. The distance center to center of each well and within each microtiter plate should be the same in all of the plates. Any deviation in movement or plate structure that positions some sample well 30 significantly off-center from line of sight to the PMT 12 causes some light from the luminescence reaction emitted from sample 10 and passing through transparent bottom 31 to become lost for striking the baffle 60 or the focussing collar 64 instead of the PMT face. Since this luminescence never reaches the PMT 12 to be counted, such loss of emitted light generates reading error. When iris 65 is smaller in diameter than well 30 as is illustrated in Fig. 6, and such is preferred, then not all of the light passing through the transparent bottom 31 from a properly centered well 30 will strike the PMT face. In short, light from the annular peripheral

edges of transparent bottom 31 is supposed to become lost. As a result, small inaccuracies in centering, either by the X-Y table and/or through minor (off center) irregularities in the sample holder plate, e.g., the ± 0.04 mm alluded to above will not generate reading error, until and unless the deviation from strict concentricity of the line of sight well 30 with PMT 12 exceeds the narrowed view path formed by aperture 65 in the iris. To repeat, an iris aperture 65 of diameter smaller than the internal diameter of well 30 is preferred.

As an aside, it is noteworthy that the home position of Table 110, the position to which X, Y mechanism 18 retreats at the end of a batch and/or hatch 102 is opened, extends Table 110 and the plates 16 thereon toward the front face of housing 100 for best access to plates 16.

Placement of sample holder plates 16 into a close fitting space on Table 110 beneath shelf 29 is not difficult. The sample holder plates are small enough for easy manipulation. The luminometer 5 is a conveniently sized unit being, for example, about 15 by 18 by 20 inches.

Some special advantages of the luminometer of this invention over like prior art devices may be made apparent by the following discussion of how the above-described components in the luminometer interact.

Self-evidently, the opaque preferably black members, i.e., sample holder plate body 35, PMT baffle 60 and focussing collar 64, serve to shield the photo sensitive face of PMT 12 from direct and reflected light. The recess 40 beneath well 30 is intended to prevent diffusion of emitted light from any well 30 in a lateral direction.

A direct line of sight path for transmission of luminescence generated in sample 10 to PMT 12 is provided by the transparent bottom 31 of each sample well 30 through the iris 65 of the focussing collar 64. No other potential

light source has such a direct path. However, prevention of any and all light leakage to the PMT face is important for best sensitivity and accuracy of the luminometer 5. Thus, if hatch 102 and/or the housing 100 itself is not
5 completely light-tight, serious reading errors will be introduced. It is important also that afterglow light from adjacent wells (out of line of sight) should be precluded from striking PMT 12. Afterglow light, sometimes preglow light as well is present. The usual luminescence reactions
10 used for assay purposes generates light for a finite period of time and the generation of light which is not an instantaneous on-off system increases to a peak level then declines and ceases.

If the luminescence assays are carried out in reasonably
15 rapid succession, more than one sample well 30 might well be emitting light while an assay is under way. Of course, conduct of each successive assay might be delayed until full extinction of the afterglow from all previous assay measurements had occurred but a lengthy time lapse between
20 successive luminescence tests is not desirable. Conduct of assays in rapid succession is preferred. In consequence, the structure of the luminometer of this invention as a whole and of sample holder plate 16 as well is adapted to allow only light from the line of sight sample well 30 to
25 reach the PMT face, preventing afterglow and/or preglow luminescence light from reaching the photosensitive PMT face.

Since a direct path for transmission of light to the PMT face is allowed only from the bottom of the sample well
30 containing the sample being assayed, light from all other wells can reach the PMT face only through reflection. Use of opaque light absorbent materials used for plate body 35, for baffle plate 60, and for collar 64 reduces reflection of light. Mention has already been made that in preferred

practice, the plate body 35, the PMT baffle and the focussing collar are black (to better absorb incident light). The inside face(s) of hatch 102 and of housing 100 also may be black.

- 5 The clearance distances above table 110 between the top of sample holder plate(s) 16 and stationary shelf 29 and the clearance beneath bottom of sample holder plate(s) 16 and the stationary baffle plate 60 are made as small as possible. The recessed location sample wells 30 and of the
10 PMT face as discussed above reduce further reduces light leakage to the PMT face.

Provision of the recess 40 beneath each sample well 30 and location of the PMT face beneath the focussing collar 64 behind the iris eliminate any line of sight passage to
15 PMT 12 from all sample wells, save from the one line of sight well 30 being assayed. Thus, afterglow from any other well 30 is never in any line of sight with the PMT face, even most obliquely. Afterglow light is directed down onto baffle 60 and obliquely from the well 30 but not
20 laterally. The recess 40 prevents lateral transmission of afterglow light. Then the recessed location of the PMT 12 face would keep any lateral light from striking the PMT face in any event. Accordingly, afterglow light from one or more wells can generate only some level of diffused
25 lighting in the space between sample holder plate 30 and the baffle 60. The black surface of plate body 35, of baffle 60, and of the focussing collar 64 absorb light, attenuating the afterglow so that even diffused (reflected) light cannot strike the PMT face.

- 30 The space overhead of the microtiter plate also contains diffused light. Some emitted light from the sample being assayed and afterglow from the previously assayed sample(s) is directed upward into this space. The black surface of the plate body 35 and of the upper shelf 29 and the inside

of housing 100 attenuate any such afterglow so that afterglow light in the overhead space is not believed to generate false PMT readings.

As an aside, it should be appreciated that the foregoing
5 discussion of afterglow light applies as much to the initial luminescence or preglow when an assay reaction involves delay and appropriately injector 14 is not in line of sight with PMT 12.

An (optional) expedient contemplated herein to increase
10 the quantity of signal light transmitted by a sample being assayed is to provide a reflector 126 in line with PMT 12, e.g., around injector nozzle 13. Reflector 126 acts to reflect back the light emitted upwardly from the sample being assayed. In addition, the reflector shields the line
15 of sight well from reflected afterglow light from adjacent wells.

It may be noted that the combination of a black plate body 35 and optically clear cups (for wells 30) may not always be desired. Although black is strongly preferred
20 for body 35, the side wall of wells 30 need not be transparent. An alternative construction contemplated by the inventors hereof employs a white or other reflective coating, e.g., mirror coating, on the side wall of each well 30. A comparative study between wells with
25 transparent side walls and wells with a white (reflective) side wall demonstrated that wells with the reflective side wall transmitted nearly double the luminescence signal.

CLAIMS

1. A luminometer comprising:

- a) a movable table adapted for positioning thereon of at least one sample holder plate characterized by having a multiplicity of sample wells in rows and columns and by each sample well having a transparent bottom wall;
- b) at least one stationary reagent injector nozzle positioned above said table;
- c) at least one stationary light detector positioned beneath said table, said reagent injector nozzle and light detector being in appropriate registry for a luminescence assay;
- d) an X-Y movement operatively connected to said table to move said table and a sample holder plate positioned on said table so that one by one every sample well can be in a line of sight assay registry with said light detector;
- e) means for preventing light emitted from any sample well not in line of sight assay registry with said light detector from impinging therein; and
- f) means for generating and displaying and/or recording a signal that is in a predetermined proportion to the intensity of the detected luminescence as the output signal assay results from said light detector.

2. The luminometer of Claim 1 further comprising a baffle plate between said table and said light detector, said baffle plate being provided with an aperture therein wherein said light detector is disposed, the baffle plate and table being located one relative to the other so that at each assay registry only a single sample well on a sample holder plate positioned on said table can be in line of sight to said light detector.

3. The luminometer of Claim 2 further comprising an iris located in said aperture on the baffle plate, the iris being located between light detector and table, said iris

limiting the maximum width line of sight path from a sample well to said light detector to less than the diameter of the sample well.

4. The luminometer of Claim 1 further comprising a reflector above said sample holder in line of sight above said light detector, whereby luminescence emitted up from the sample well on said plate that is in line of sight to said light detector will be reflected back down through said sample.

5. The luminometer of Claim 1 wherein the light detector comprises a photo multiplier tube.

6. The luminometer of Claim 1 wherein the means for preventing light from any sample well not in line of sight with said light detector from impinging thereon comprises a black surface absorptive of light on the internal surface of said luminometer.

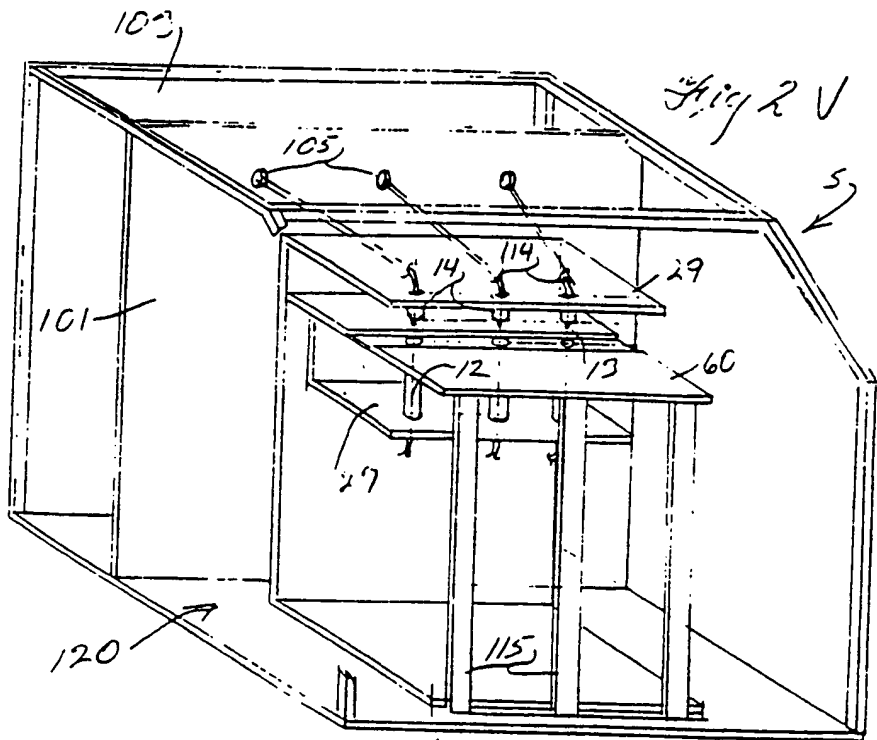
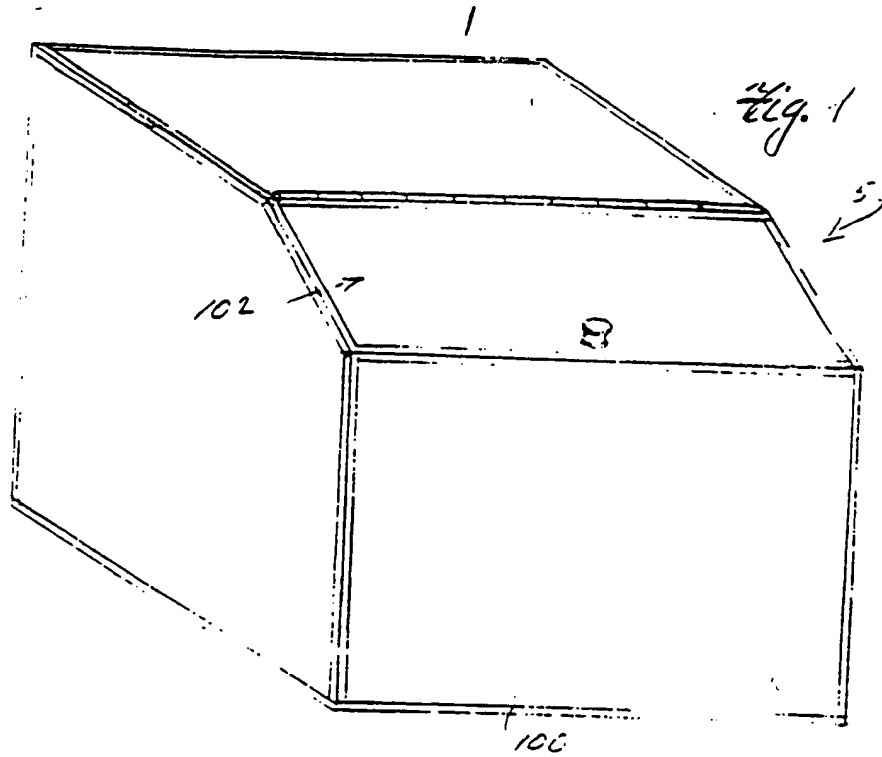
7. The luminometer wherein said movable table is adapted to position thereon three sample holder plates; and wherein three nozzles and three light detectors are present, whereby three samples may be assayed at the same time.

8. A sample holder for a luminometer assay comprising a body member having therein a multiplicity of pass through cylindrical apertures in an array of columns and rows, each aperture being plugged by a cup shaped sample well open at the top for introduction of sample therein and closed at the bottom by a transparent bottom wall, the transparent bottom wall being recessed within the aperture, the side wall of said sample well being transparent or reflective.

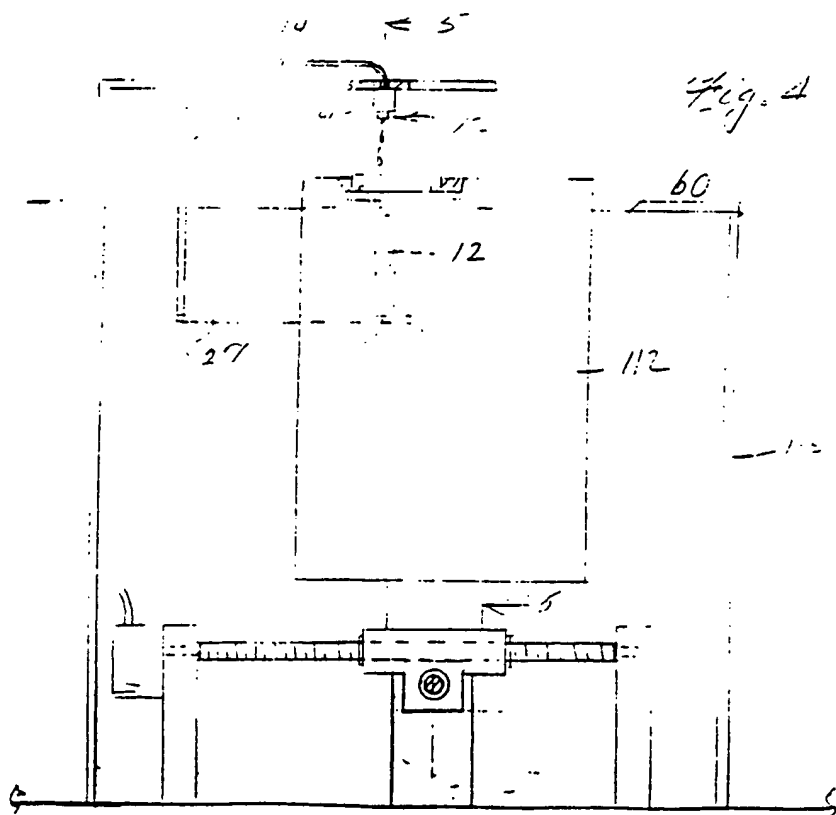
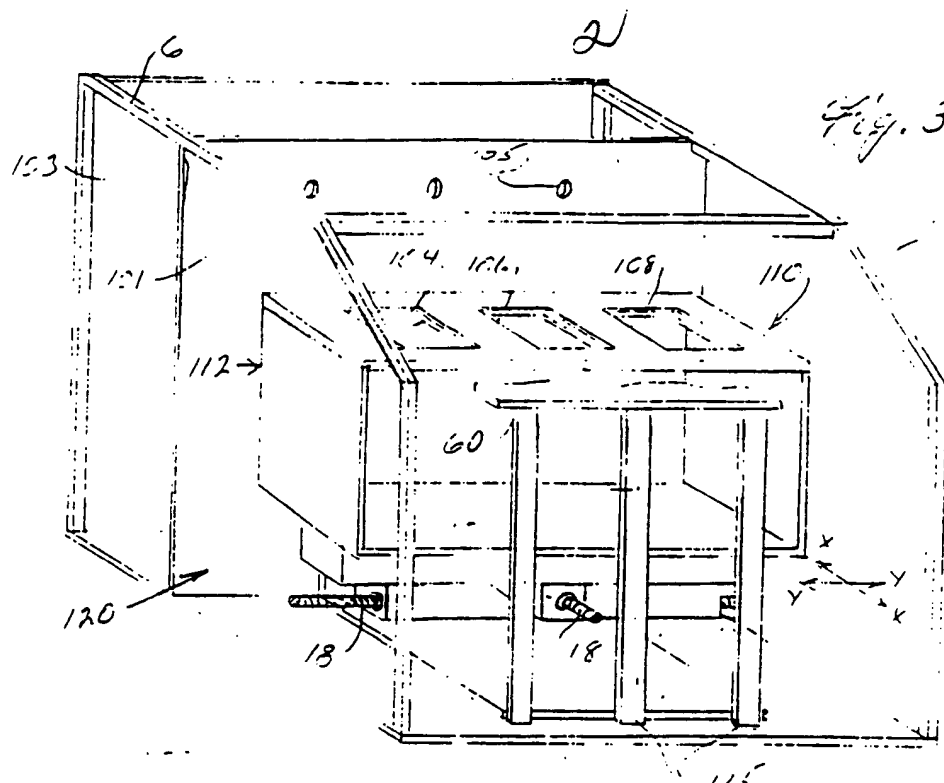
9. A sample holder as in Claim 7 dimensioned to be the same dimensions as the state of the art microtiter plates.

10. A sample holder as in Claim 7 wherein the body member thereof is black and the sample wells contain reflective side walls.

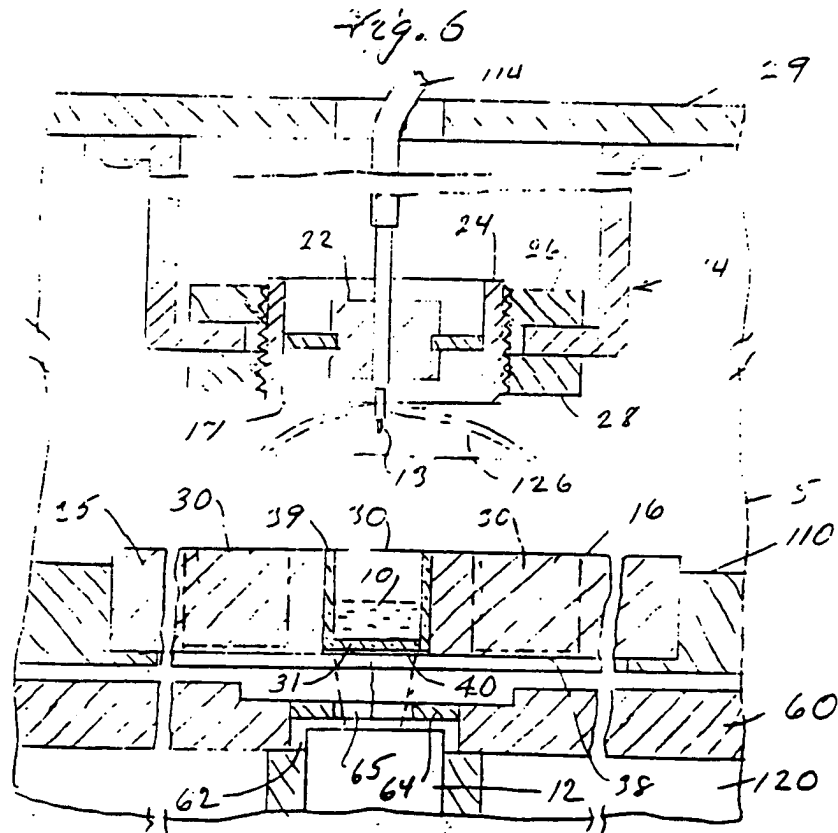
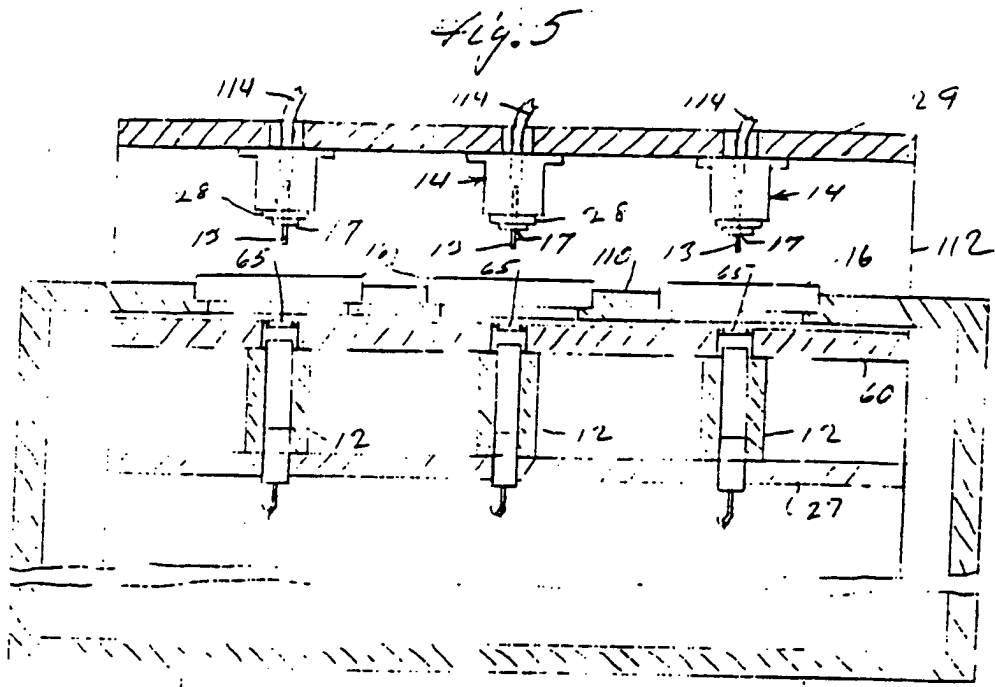
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INTERNATIONAL SEARCH REPORT

International Application No. PCT/US90/05238

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all) ¹		
According to International Patent Classification (IPC) or to both National Classification and IPC IPC 5): G01N 21/00, 21/76, 1/10 U.S. Cl.: 422/82.05, 82.08; 436/172, 809; 356/246, 440		
II. FIELDS SEARCHED		
Minimum Documentation Searched ⁴		
Classification System	Classification Symbols	
US CL.	422/52, 82.05, 82.08, 104; 436/172, 809; 206/569; 356/246, 434, 436, 440	
Documentation Searched other than Minimum Documentation to the extent that such documents are included in the fields searched ⁵		
III. DOCUMENTS CONSIDERED TO BE RELEVANT ¹⁴		
Category ⁶	Citation of Document, ¹⁶ with indication, where appropriate, of the relevant passages ¹⁷	Relevant to Claim No. ¹⁸
Y	US, A, 4,772,453, (Lisenbee) 20 September 1988, see col. 4, lines 3-7, 30-38, 39-48, 57-58, col. 5, lines 1-4, col. 8, lines 65-68, col. 15, claim 15.	1-7
Y	EP, A, 0,217,632, (Chow et al.), 08 April 1987, see col. 6, lines 18-23, col. 7, lines 6-13, 16-26.	1-7
Y	US, A, 4,431,307, (Suovanen), 14 February 1984, see col. 1, lines 30-45.	6,8-10
A	US, A, 4,818,883 (Anderson et al.) 04 April 1989, see entire document.	1-7
A	US, A, 4,755,055, (Johnson et al.) 05 July 1988, see entire document.	1-7
<p>* Special categories of cited documents: ¹⁵</p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step</p> <p>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.</p> <p>"Δ" document member of the same patent family</p>		
IV. CERTIFICATION		
Date of the Actual Completion of the International Search ¹	Date of Mailing of this International Search Report ¹	
16 January 1991	07 FEB 1991	
International Searching Authority ¹	Signature of Authorized Officer ²	
ISA/US	Theresa A. Trembley	

Form PCT/ISA/210 (second sheet) (May 1986)

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)		
Category *	Citation of Document, ¹⁶ with indication, where appropriate, of the relevant passages ¹⁷	Relevant to Claim No ¹⁸
A	US, A, 4,115,010, (McAleer et al.) 19 September 1978, see entire document.	1-7
A	US, A, 4,730,921 (Klein et al.) 15 March 1988, see entire document.	1-7
A	US, A, 4,826,660, (Smith et al.), 02 May 1989, see entire document.	1-7
A	UK, A, 2,211,607, (Wilson et al.), 05 July 1989, see entire document.	1-7
Y	US, A, 4,735,778, (Naruyama et al.), 05 April 1988, see col. 2, lines 62-65, col. 3, lines 14-16.	8-10
Y	US, A, 4,684,250, (Kukka et al.) 04 August 1987, see col. 1, lines 31-40.	8-10
Y	US, A, 4,828,386 (Matkovich et al.) 09 May 1989, see col. 1, lines 27-28, 42-45, col. 3, lines 34-47.	8-9
Y, P	US, A, 4,956,150, (Henry) 11 September 1990, see col. 1, lines 46-50.	8-10

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FURTHER INFORMATION CONTINUED FROM THE SECOND SHEET

For the purposes of this search report: Claim 7 appears to be a dependent claim, however fails to refer back to a prior claim, assumed dependency is to claim 1.

It was assumed that claims 9 and 10 were meant to be referred to claim 8.

V. ☐ OBSERVATIONS WHERE CERTAIN CLAIMS WERE FOUND UNSEARCHABLE¹

This international search report has not been established in respect of certain claims under Article 17(2) (a) for the following reasons:

1. ☐ Claim numbers _____, because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claim numbers _____, because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out¹, specifically:

3. ☐ Claim numbers _____, because they are dependent claims not drafted in accordance with the second and third sentences of PCT Rule 6.4(a).

VI. ☒ OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING²

This International Searching Authority found multiple inventions in this international application as follows:

I. Claims 1-7 drawn to a luminometer, classified in class 422 subclass 82.05.

II. Claims 8-10 drawn to a sample holder, classified in class 422 subclass 104.

1. ☒ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims of the international application.
2. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims of the international application for which fees were paid, specifically claims:

3. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claim numbers:

4. ☐ As all searchable claims could be searched without effort justifying an additional fee, the International Searching Authority did not invite payment of any additional fee.

Remark on Protest

- ☐ The additional search fees were accompanied by applicant's protest.
- ☒ No protest accompanied the payment of additional search fees.